

The Extracellular Domain of p75^{NTR} Is Necessary to Inhibit Neurotrophin-3 Signaling through TrkA*

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The TrkA receptor is activated primarily by nerve growth factor (NGF), but it can also be activated by high concentrations of neurotrophin 3 (NT-3). The pan-neurotrophin receptor p75^{NTR} strongly inhibits activation of TrkA by NT-3 but not by NGF. To examine the role of p75^{NTR} in regulating the specificity of TrkA signaling, we expressed both receptors in *Xenopus* oocytes. Application of NGF or NT-3 to oocytes expressing TrkA alone resulted in efflux of ⁴⁵Ca²⁺ by a phospholipase C- γ -dependent pathway. Coexpression of p75^{NTR} with TrkA inhibited ⁴⁵Ca²⁺ efflux in response to NT-3 but not NGF. The inhibitory effect on NT-3 activation of TrkA increased with increasing expression of p75^{NTR}. Coexpression of a truncated p75^{NTR} receptor lacking all but the first 9 amino acids of the cytoplasmic domain inhibited NT-3 stimulation of ⁴⁵Ca²⁺ efflux, whereas coexpression of an epidermal growth factor receptor/p75^{NTR} chimera (extracellular domain of epidermal growth factor receptor with transmembrane and cytoplasmic domains of p75^{NTR}) did not inhibit NT-3 signaling through TrkA. These studies demonstrated that the extracellular domain of p75^{NTR} was necessary to inhibit NT-3 signaling through TrkA. Remarkably, p75^{NTR} binding to NT-3 was not required to prevent signaling through TrkA, since occupying p75^{NTR} with brain-derived neurotrophic factor or anti-p75 antibody (REX) did not rescue the ability of NT-3 to activate ⁴⁵Ca²⁺ efflux. These data suggested a physical association between TrkA and p75^{NTR}. Documenting this physical interaction, we showed that p75^{NTR} and TrkA could be coimmunoprecipitated from *Xenopus* oocytes. Our results suggest that the interaction of these two receptors on the cell surface mediated the inhibition of NT-3-activated signaling through TrkA.

Nerve growth factor (NGF),¹ brain-derived neurotrophic factor (BDNF), neurotrophin-4 (NT4), and neurotrophin-3 (NT-3) initiate their actions by binding to Trk receptors and p75^{NTR} (1). NGF binds to TrkA; BDNF and NT-4 bind to TrkB, and NT-3 binds to TrkC. NT-3 also binds TrkA (and TrkB), although with significantly lower affinity (1), and can activate TrkA signaling and cellular responses (2–7). *In vivo*, NT-3 signaling through other Trk receptors is required for nervous system development, since NT3^{–/–} mice exhibit significantly greater neuronal loss than TrkC^{–/–} mice (8). Recently, NT-3 has been shown to be required for the survival of TrkA-expressing postmitotic sympathetic neurons (9) and sensory neurons (10) *in vivo*.

p75^{NTR} also binds each of the neurotrophins (11) and can signal independently of TrkA in response to neurotrophin stimulation (12–14). In addition, p75^{NTR} regulates Trk receptor signaling. p75^{NTR} potentiates NGF activation of TrkA at low ligand concentrations (15, 16) and collaborates with TrkA to form high affinity binding sites (17–19). Furthermore, the signaling pathways initiated by p75^{NTR} block Trk receptor signaling in certain contexts (20, 21) and synergize with Trk receptor signaling in other contexts (22). These opposing actions suggest a complex interaction between p75^{NTR} and Trk receptors in regulating neurotrophin signaling.

NT-3 does not activate TrkA signaling in PC12 cells (4, 23), unless p75^{NTR} levels are reduced (23), suggesting that p75^{NTR} directly suppresses the ability of TrkA to respond to NT-3 (4). In support of this hypothesis, isolated sympathetic neurons from p75^{NTR}^{–/–} mice are more responsive to NT-3 in culture than are wild type sympathetic neurons (24). These data suggest that p75^{NTR} can prevent NT-3 signaling through TrkA, but the molecular nature of this interaction remains to be characterized.

We used a *Xenopus* oocyte microinjection assay to study NGF and NT-3 signaling through TrkA and to assess the role of p75^{NTR} in regulating this signaling. We showed that p75^{NTR} prevented the signaling of NT-3 through TrkA but not that of NGF. We demonstrated that the extracellular domain of p75^{NTR}, but not the cytoplasmic domain, was necessary to inhibit NT-3 signaling through TrkA. Furthermore, p75^{NTR} binding to NT-3 was not required to inhibit NT-3 activation of TrkA. Finally, we showed that p75^{NTR} and TrkA could be coimmunoprecipitated from *Xenopus* oocytes, suggesting that

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¹ The abbreviations used are: NGF, nerve growth factor; NT-3, neurotrophin-3; BDNF, brain-derived neurotrophic factor; NT-4, neurotrophin-4; p75^{NTR}, p75 neurotrophin receptor; EGFR, epidermal growth factor receptor; PAGE, polyacrylamide gel electrophoresis; PCR, polymerase chain reaction.

interaction of these receptors mediates the inhibition of NT-3 signaling through TrkA.

EXPERIMENTAL PROCEDURES

Growth Factors and Antibodies—Purified recombinant human neurotrophins were obtained from Amgen Inc. (Thousand Oaks, CA) and Genentech Corp. (South San Francisco, CA). Rabbit polyclonal anti-p75^{NTR} antibody was raised against the extracellular domain of rat p75^{NTR} (25). Rabbit polyclonal antibody raised against the cytoplasmic domain of p75^{NTR} was purchased from Promega (Madison WI). Rabbit polyclonal antibody (26) was raised to the C terminus of human TrkA. M2 monoclonal anti-FLAG antibody was purchased from Stratagene (La Jolla, CA), and monoclonal anti-EGFR antibody (LA22) was purchased from Upstate Biotechnology, Inc. (Lake Placid, NY).

cDNA Constructs and Plasmid Generation—cDNAs encoding the full length rat TrkA and rat p75^{NTR} receptors were subcloned into PGEMHE vector, which contains 5' and 3' *Xenopus* beta-globin untranslated sequences for RNA stability (gift from Dr. E. R. Liman) (27). Truncated p75^{NTR} receptor was generated from the rat p75^{NTR} cDNA using PCR with forward primer 5' GAA TTC ATG AGG AGG GCA GGT GCT GCC and reverse primer 5' CAC GGG TCT AGA CTA GGC GCC TTG TTT ATT TTG TTT GCA GCT G to amplify the extracellular and transmembrane domains, and the first nine amino acids of the cytoplasmic domain, flanked by a 5' *EcoRI* site and a 3' *XbaI* site. The amplified product was subcloned into PGEMHE. All constructs used in these experiments were verified by sequencing.

A chimeric EGF receptor/p75^{NTR} cDNA encoding the extracellular domain of the human EGF receptor with the transmembrane and cytoplasmic domains of human p75^{NTR} (gift of Dr. Moses Chao) (28) was subcloned from pCMVEN10 into the PGEMHE vector. This was done using PCR with forward primer 5' GCG CGC GTC GAC GCG ATG CGA CCC TCC GGG ACG GCC and reverse primer 5' GCG CGC TCT AGA GGC TCA CAC CGG GGA TGA GGC AGT GG to amplify the chimeric EGFR/p75^{NTR} construct flanked by a 5' *Sall* site and a 3' *XbaI* site. The amplified product was digested with *Sall* and *XbaI* and subcloned into PGEMHE.

The C-terminal FLAG-tagged TrkA receptor was generated by PCR using forward primer 5' CA GGG ACT AGT GGT CAA GAT GAT TGG A and reverse primer 5' CTT ATC ATC ATC ATC CTT GTA ATC GCC CAG AAC GTC CAG G to amplify a fragment encoding the last 405 base pairs of rat TrkA receptor coding sequence (minus the stop codon) including the 5' *SpeI* site (part of the coding sequence) and the first FLAG epitope on the 3' end. This product was then used as a template for a second PCR using the same forward primer and a reverse primer encoding the second FLAG tag followed by a stop codon and an *XhoI* restriction site 5' CTG CTC GAG CTA CTT ATC ATC ATC CTT GTA ATC CTT ATC ATC ATC. The amplified product was digested with *SpeI* and *XhoI* and subcloned into similarly digested full-length TrkA in PGEMHE.

In Vitro Transcription of cRNA and Expression in *Xenopus* Oocytes—DNA templates were linearized and transcribed *in vitro* using the T7 RNA polymerase promoter with mMessage *in vitro* transcription kit from Ambion (Austin, TX). Transcripts were purified using an RNeasy kit purchased from Qiagen Inc. (Valencia, CA) and then analyzed on agarose gels and quantified by spectrophotometry (as well as by densitometric comparison of bands to known RNA standards).

***Xenopus laevis* and Preparation of Oocytes**—Mature *Xenopus* oocytes (Dumont stage V–VI) were harvested and defolliculated enzymatically with 1–2 mg/ml collagenase (type I or II) for 2–3 h (Worthington). Oocytes were maintained at 17–18 °C in ND96 (96 mM NaCl, 2 mM KCl, 1.8 mM CaCl₂, 1 mM MgCl₂, 5 mM HEPES, pH 7.5) plus 1 µg/ml ciprofloxacin, 6 µg/ml ceftazidime, and 1 µg/ml gentamicin.

⁴⁵Ca²⁺ Efflux Assays—For calcium efflux assays, oocytes were injected with a total of 0.2 ng of cRNA per oocyte. When cRNA encoding a single receptor was injected, 0.1 ng of GFP cRNA was coinjected to equalize the total amount of injected cRNA. Efflux experiments were conducted according to methods published previously (29, 30). In brief, 2 days after injection, oocytes were incubated in ⁴⁵Ca²⁺ (from Amersham Pharmacia Biotech), 100 mCi/ml for 2.5 h in calcium-free ND96. Groups of 8 (or 4) oocytes were washed and transferred to 24-well dishes with 0.5 ml of ND96 per well. Medium was collected and replaced every 10 min, and radioactivity was measured by liquid scintillation counting. After stabilization of background ⁴⁵Ca²⁺ efflux, 100 ng/ml either NT-3 or NGF was added to each group of oocytes. Data were collected from each of the experimental conditions by using 4–8 oocytes per condition (same number of oocytes in each experiment), with each condition run in triplicate for each experiment. The initial NT-3 dose-response curve

(Fig. 2) was run in duplicate. Each experiment was repeated two or three times, using oocytes from two or three different frogs. Therefore, every data point for each condition in one experiment consisted of the average of three measurements obtained from two or three groups of four to eight oocytes. Signaling curves were graphed using Microsoft Excel, and mean and standard errors of measure were determined using Microsoft Excel. For analysis of dose-response curve, mean peak counts per min were compared using analysis of variance methods and allowing for unequal variances. The Tukey-Fisher criterion was used to compute post hoc *t* statistics and their corresponding *p* values. Similar results were obtained using a log scale transformation of the data where equal variances could be assumed.

Immunoprecipitations and Immunoblotting—For analysis of TrkA, p75^{NTR}, and truncated p75^{NTR} receptors, *Xenopus* oocytes were injected with each cRNA (1–5 ng per oocyte of each cRNA). Thirty six hours post-injection, 25 oocytes from each condition were manually homogenized in 250 µl of ice-cold solubilization buffer (65 mM Tris-HCl, pH 7.5, 50 mM NaCl, 5 mM EDTA, 2% Triton X-100, 1% Nonidet P-40, 0.5% sodium deoxycholate, 0.1% SDS, 1 mM phenylmethylsulfonyl fluoride, 5 µg/ml aprotinin, 5 µg/ml leupeptin, 1 mM sodium orthovanadate, and 10 mM sodium fluoride). Lysates were incubated at 4 °C for 1 h, and the soluble phase was partitioned from the insoluble and lipid phases by three rounds of centrifugation at 18,370 × *g* for 5 min at 4 °C. 250 µl of 2× IP buffer (solubilization buffer with 10 mg/ml of bovine serum albumin) was added to each sample and pre-cleared with protein A- or protein G-Sepharose beads for 1 h. TrkA receptors were immunoprecipitated with the anti-FLAG monoclonal antibody (M2), and P75^{NTR} and p75⁻ receptors were precipitated using wheat germ lectin-agarose beads (Amersham Pharmacia Biotech). Precipitated samples were washed four times with 1× solubilization buffer, after which precipitated proteins were removed from the beads by boiling for 5 min in 3× Laemmli sample buffer, and SDS-PAGE was performed on 7.5% polyacrylamide gels. After transfer to nitrocellulose membranes (Hybond, Amersham Pharmacia Biotech), samples were blocked in TBST with nonfat dried milk and incubated with primary antibodies against the FLAG epitope of TrkA (Stratagene, La Jolla CA), the extracellular domain of p75^{NTR} (REX), or the cytoplasmic domain of p75^{NTR} (Promega, Madison, WI) overnight at 4 °C. After washing, blots were incubated with horseradish peroxidase-conjugated goat anti-mouse (Promega, Madison, WI or New England Biolabs, Beverly, MA) or horseradish peroxidase-conjugated goat anti-rabbit (Promega, Madison, WI or New England Biolabs, Beverly, MA) at a dilution of 1:5000 for 1–2 h at room temperature. After washing, bands were visualized by incubating with Supersignal (Pierce) or ECL (Amersham Pharmacia Biotech), followed by exposure to Kodak BioMax film (Eastman Kodak Co.).

For analysis of the chimeric EGFR/p75^{NTR} receptor, *Xenopus* oocytes were injected with 5 ng per oocyte of EGFR/p75^{NTR} cRNA. Thirty six hours post-injection, 25 oocytes were manually homogenized in ice-cold homogenization buffer (65 mM Tris-HCl, pH 7.5, 10 mM EDTA, 1 mM phenylmethylsulfonyl fluoride, 5 µg/ml aprotinin, 5 µg/ml leupeptin, 1 mM sodium orthovanadate, and 10 mM sodium fluoride), and lysates were centrifuged at 735 × *g* for 5 min at 4 °C to separate a heavy fraction (containing the plasma membrane and cortical granules) from the cytoplasmic fraction. The heavy fraction was solubilized in 250 µl of RIPA buffer (with 2.5% Triton X-100) for 1 h at 4 °C and centrifuged at 18,370 × *g* for 20 min at 4 °C. Samples were pre-cleared with protein G-Sepharose beads. EGFR/p75^{NTR} receptors were immunoprecipitated with anti-EGFR monoclonal antibody (clone LA22, Upstate Biotechnology, Inc., Lake Placid, NY) overnight at 4 °C. Precipitated samples were washed four times with 1× solubilization buffer, after which precipitated proteins were removed from the beads by boiling for 5 min in 3× Laemmli sample buffer, and SDS-PAGE was performed on 7.5% polyacrylamide gels. After transfer to nitrocellulose membranes (Hybond, Amersham Pharmacia Biotech), samples were blocked in TBST with nonfat dried milk and incubated with anti-EGFR antibody.

For coimmunoprecipitation experiments, equal amounts of cRNA for FLAG-tagged TrkA and p75^{NTR} (5 ng of each cRNA) were microinjected into each oocyte. After 48 h, oocytes were incubated with NT-3 (100 ng/ml), NGF (100 ng/ml), or no neurotrophin for 20 min at room temperature, after which they were placed on ice. Oocytes were manually homogenized in ice-cold homogenization buffer, and samples were centrifuged at 735 × *g* for 5 min at 4 °C. The pellet (containing the membrane) was solubilized in 500 µl of a buffer containing 130 mM Tris-HCl, pH 7.5, 150 mM NaCl, 2% Triton X-100, and 10 mM EDTA, 1 mM phenylmethylsulfonyl fluoride, 1 µg/ml aprotinin, 1 µg/ml leupeptin, and 1 mM sodium orthovanadate. The cytoplasmic fraction was solubilized by adding a 2× concentration of this solubilization buffer to

the 250 μ l of supernatant. Fractions were solubilized for 15 min at 37 °C and centrifuged at 18,370 $\times g$ for 20 min at 4 °C. Samples were pre-cleared with protein A/G-Sepharose beads and immunoprecipitated with anti-p75^{NTR} (REX) (25). After SDS-PAGE and transfer, nitrocellulose membranes (Hybond, Amersham Pharmacia Biotech) were blocked in TBST with nonfat dried milk and incubated with primary antibodies against p75^{NTR} extracellular domain (REX) (25) or anti-FLAG M2 monoclonal antibody (Stratagene, La Jolla, CA).

RESULTS

Expression of TrkA and p75^{NTR} in *Xenopus* Oocytes—The *Xenopus* oocyte ⁴⁵Ca²⁺ efflux assay has been used to quantify receptor activation and signal transduction (29, 30). Mature oocytes are capable of translating injected cRNA, targeting proteins to correct subcellular locations, and inserting receptors into the plasma membrane (31). Because the phospholipase C- γ pathway is conserved in oocytes, activation of receptor tyrosine kinases, such as Trk receptors, stimulates phospholipase C- γ and phosphatidylinositol turnover and stimulates intracellular calcium release (32, 33). The increased intracellular calcium activates a chloride current in *Xenopus* oocytes and causes cytoplasmic calcium to be released into the extracellular medium (32, 34, 35). Receptor activation and signal transduction through the phospholipase C- γ pathway can therefore be quantified by using a ⁴⁵Ca²⁺ efflux assay (29, 30). For this reason, we chose to express TrkA (with or without p75^{NTR}) in *Xenopus* oocytes to quantitatively study NT3 activation of TrkA and the role of p75^{NTR} in regulating it.

To determine whether TrkA and p75^{NTR} protein expression correlate with the amount of each cRNA injected, we microinjected *Xenopus* oocytes with 0, 1, 2.5, or 5 ng of TrkA per oocyte and determined the level of TrkA protein expressed. There was a proportional increase in the amount of TrkA receptor protein expressed by oocytes in response to increasing amounts of injected TrkA cRNA (Fig. 1A). Similarly, oocytes injected with 0, 1, or 2.5 ng of p75^{NTR} cRNA demonstrated that p75^{NTR} protein expression correlated with the amount of p75^{NTR} cRNA injected (Fig. 1B). To determine whether coexpression of p75^{NTR} interferes with TrkA receptor expression, oocytes were coinjected with TrkA cRNA (1 ng per oocyte) and increasing amounts of p75^{NTR} cRNA (0, 1, or 2.5 ng per oocyte). Injection of increasing amounts of p75^{NTR} cRNA resulted in increasing p75^{NTR} expression; however, the amount of TrkA expressed was unaffected by p75^{NTR} coexpression (Fig. 1C). Thus, the amount of TrkA and p75^{NTR} protein expressed correlated with the amount of each cRNA injected, similar to other proteins exogenously expressed in *Xenopus* oocytes (29, 30, 36).

NT-3 Activates Signaling through TrkA Receptors—Having determined that TrkA and p75^{NTR} receptor expression correlate with the amount of each cRNA injected, we next studied the signaling of TrkA-expressing *Xenopus* oocytes in response to a range of NT-3 concentrations. NT-3 activated TrkA signaling through the phospholipase C- γ pathway in a dose-dependent fashion (Fig. 2). Peak response was seen 30 min after addition of 10 ng/ml NT-3, 20 min after addition of 25 or 50 ng/ml of NT-3 and 10 min after addition of 100, 200, or 300 ng/ml NT-3 (data not shown). The peak response to NT-3 plateaued at about 100 ng/ml, suggesting that 100 ng/ml was functionally saturating. Similar results were obtained using a log scale transformation of the data where equal variances could be assumed. In addition, differences in mean peak signaling were compared using one-way analysis of variance. There were no statistically significant mean differences in NT-3 signaling at 100, 200, and 300 ng/ml ($p > 0.52$ or larger for the three comparisons). However, the mean response at 100 ng/ml was significantly different from the response at 50 ng/ml ($p = 0.002$). All subsequent experiments were done using 100 or 200 ng/ml of NT-3.

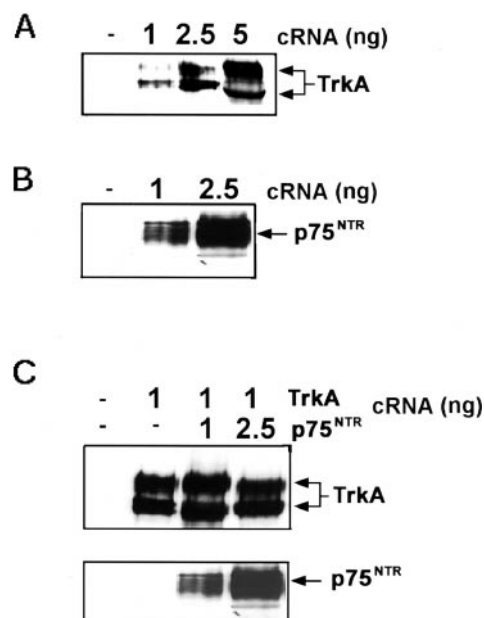


FIG. 1. TrkA and p75^{NTR} receptor expression are highly correlated with the amount of cRNA injected into *Xenopus* oocytes, and coexpression of p75^{NTR} does not interfere with TrkA receptor expression. A, the amount of TrkA receptor protein expressed correlates with the amount of TrkA receptor cRNA injected. *Xenopus* oocytes were injected with 0, 1, 2.5, or 5 ng of TrkA cRNA, and the level of TrkA protein expressed was determined by immunoprecipitation followed by immunoblotting. There is a proportional increase in the amount of TrkA receptor, 140- and 110-kDa doublet, expressed in response to increasing amounts of TrkA cRNA injected. B, the amount of p75^{NTR} protein expressed correlates with the amount of p75^{NTR} cRNA injected. Oocytes were injected with 0, 1, or 2.5 ng of p75^{NTR} cRNA, and p75^{NTR} protein expression was determined by precipitation with wheat germ lectin-agarose followed by immunoblotting. A proportional increase in p75^{NTR} protein, 75–80 kDa, is seen with increasing p75^{NTR} cRNA injection. C, increasing p75^{NTR} expression does not alter the level of TrkA receptor expressed. Oocytes were injected with 1 ng of TrkA and increasing amounts of p75^{NTR} (0, 1, and 2.5 ng per oocyte). p75^{NTR} expression is noted to increase proportionally; however, TrkA expression is not altered by p75^{NTR} coexpression.

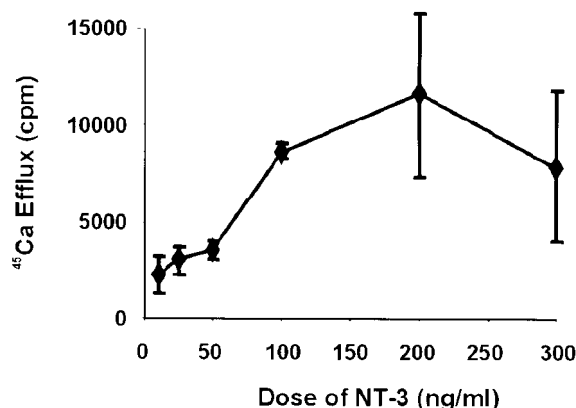


FIG. 2. Dose-response curve of ⁴⁵Ca efflux after NT-3 application to TrkA-expressing oocytes. Oocytes expressing TrkA (0.1 ng of cRNA/oocyte) were loaded with ⁴⁵Ca²⁺ and washed (see “Experimental Procedures”). After ⁴⁵Ca²⁺ efflux levels stabilized, NT-3 was added (at time 0) to the medium at final concentrations ranging from 10 to 300 ng/ml. Peak mean counts/min were plotted against the dose of NT-3. Peak mean response to NT-3 was seen after 30 min in response to 10 ng/ml NT-3, after 20 min for 25 and 50 ng/ml of NT-3, and 10 min after addition of 100, 200, or 300 ng/ml of NT-3 (data not shown).

p75^{NTR} Specifically Inhibited NT-3 Signaling through TrkA—To determine whether p75^{NTR} inhibited NT-3 signaling through TrkA and to assess whether this inhibition was specific to NT-3, we coexpressed TrkA and p75^{NTR} in oocytes.

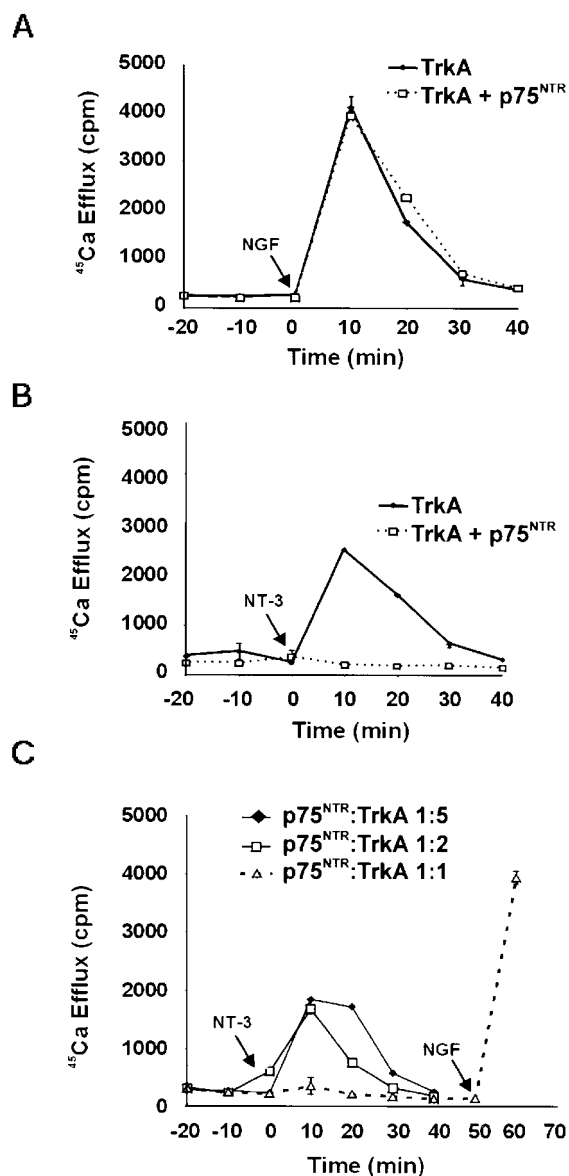


FIG. 3. p75^{NTR} inhibits NT-3 signaling through TrkA but not NGF signaling. A, oocytes expressing TrkA alone or in combination with p75^{NTR} (see “Experimental Procedures”) were loaded with ⁴⁵Ca²⁺. After stabilization of ⁴⁵Ca²⁺ efflux, 100 ng/ml NGF (A) or 100 ng/ml NT-3 (B) was added at time 0 (indicated by arrow), and calcium efflux was measured every 10 min. C, inhibitory effect of p75^{NTR} on NT-3 signaling through TrkA increases with increasing expression of p75^{NTR}. Oocytes injected with TrkA receptor cRNA (0.2 ng/oocyte) and p75^{NTR} (ranging from 0.04 to 0.2 ng/oocyte) were loaded with ⁴⁵Ca²⁺. After stabilization of ⁴⁵Ca²⁺ efflux, 100 ng/ml of NT-3 was added at time 0 (indicated by arrow), and calcium efflux was measured every 10 min. Subsequent response of oocytes to application of 100 ng/ml of NGF (arrow) demonstrates that the oocytes are capable of responding to NGF stimulation.

Equal amounts of cRNA encoding p75^{NTR} and TrkA were injected. Oocytes that expressed TrkA alone were also injected with an equal amount of green fluorescent protein (GFP) cRNA to control for total RNA injected. (GFP had no effect on the signaling response to neurotrophins, data not shown.) After loading the oocytes with ⁴⁵Ca²⁺ and allowing for stabilization of ⁴⁵Ca²⁺ efflux, 100 ng/ml of either NGF or NT-3 was added, and ⁴⁵Ca²⁺ efflux was measured every 10 min. Application of NGF or NT-3 to TrkA-expressing oocytes elicited ⁴⁵Ca²⁺ efflux. NGF signaling was unaffected by coexpression of p75^{NTR}, and p75^{NTR} had no effect on the timing or peak level of NGF signaling (Fig. 3A). In contrast, p75^{NTR} completely inhibited

NT-3 signaling through TrkA (Fig. 3B). The inhibitory effect of p75^{NTR} on NT-3 signaling through TrkA increased with increasing expression of p75^{NTR} (Fig. 3C). Remarkably, almost full potency of TrkA signaling was recovered when the p75^{NTR}:TrkA ratio was decreased from 1:1 to 1:2. These data are evidence that rather small changes in p75^{NTR} expression have marked effects on TrkA signaling. Increased expression of p75^{NTR} relative to TrkA did not impair the response of oocytes to subsequent NGF stimulation (Fig. 3C). These data demonstrate that p75^{NTR} specifically inhibits NT-3 signaling through TrkA and that the level of inhibition depends on the ratio of p75^{NTR} relative to TrkA.

The Extracellular Domain of p75^{NTR} Inhibited NT-3 Signaling through TrkA—To determine which structural domain of p75^{NTR} mediated the inhibitory effect on NT-3 signaling through TrkA, we constructed a mutant p75^{NTR} receptor with a truncated cytoplasmic domain (containing only the first nine amino acids of the cytoplasmic domain). We coexpressed this truncated p75^{NTR} with TrkA to assess its effect on NT-3 signaling. Expression of the truncated p75^{NTR} receptor was confirmed by immunoblotting (Fig. 4B), and surface expression was further confirmed by ¹²⁵I-NT-3 binding and cross-linking studies (data not shown). After stabilization of ⁴⁵Ca²⁺ efflux, 200 ng/ml of NT-3 was added, and ⁴⁵Ca²⁺ efflux was measured every 10 min. The truncated p75^{NTR} completely inhibited NT-3 signaling through TrkA (Fig. 4A). This suggested that the cytoplasmic domain of p75^{NTR} (excluding the juxtamembrane region) is not required to inhibit NT-3 signaling through TrkA. To determine whether the extracellular domain of p75^{NTR} is necessary, we coexpressed TrkA along with an EGFR/p75^{NTR} chimera, containing the extracellular domain of the EGFR fused to the transmembrane and cytoplasmic domains of p75^{NTR}. This chimeric receptor has been used to study p75^{NTR} signaling in PC12 cells and has been shown to be expressed on the cell surface (28). Expression in oocytes was confirmed by the presence of a 130-kDa band, consistent with the predicted size (28) (Fig. 4D). NT-3 signaling in *Xenopus* oocytes expressing the EGFR/p75^{NTR} receptor along with TrkA was as robust as in oocytes expressing TrkA alone (Fig. 4B). From these experiments, we conclude that the extracellular domain of p75^{NTR} is necessary for inhibition of NT-3 signaling through TrkA. We also conclude that the transmembrane and cytoplasmic domains of p75^{NTR} are not sufficient to inhibit NT-3 signaling through TrkA.

Antagonistic interactions between the signaling pathways of Trk receptors and p75^{NTR} have been described (21, 37, 38). However, our finding that the extracellular domain of p75^{NTR} inhibited NT-3 signaling, whereas most of the cytoplasmic domain was dispensable, suggests that events occurring between the receptors on the plasma membrane and between their extracellular domains are critical for inhibiting NT-3 signaling through TrkA.

p75^{NTR} Inhibited NT-3 Activation of TrkA Even When It Did Not Bind NT-3—This raised two possibilities as follows: either p75^{NTR} interacts with NT-3 in a manner that inhibits its ability to activate TrkA or p75^{NTR} interacts with TrkA to inhibit activation by NT-3. The first possibility requires p75^{NTR} binding to NT-3 to suppress signaling. Under the second possibility, NT-3 binding to p75^{NTR} is dispensable. To distinguish between these possibilities, we performed a ⁴⁵Ca²⁺ efflux assay using oocytes coexpressing both receptors, in the presence or absence of anti-p75^{NTR} antibody or excess BDNF (1 μg/ml), each of which prevents NT-3 binding to p75^{NTR}. Oocytes expressing TrkA and p75^{NTR} or TrkA alone (along with GFP) were loaded with ⁴⁵Ca²⁺ as described above. Oocytes expressing both receptors were incubated for 2 h prior to the addition of NT-3 with 1

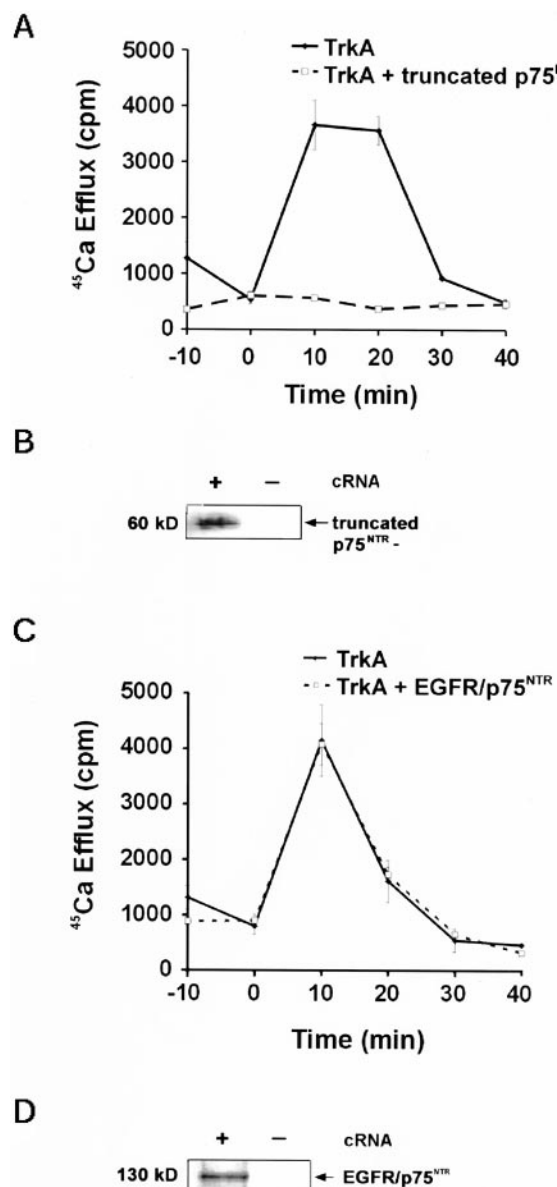


FIG. 4. The extracellular domain of p75^{NTR} is necessary to prevent NT-3 signaling through TrkA, and the transmembrane and cytoplasmic domains are not sufficient to inhibit signaling. A, a truncated mutant p75^{NTR} receptor with a minimal cytoplasmic domain (9 amino acids) inhibits NT-3 signaling through TrkA. Oocytes expressing TrkA alone in combination with truncated p75^{NTR} were loaded with ⁴⁵Ca²⁺. After stabilization of ⁴⁵Ca²⁺ efflux levels, 200 ng/ml of NT-3 was added to the medium (indicated by arrow), and ⁴⁵Ca²⁺ efflux was measured every 10 min. B, oocytes injected with p75^{NTR}-cRNA express the truncated p75^{NTR} receptor protein. Lysates from oocytes injected with p75^{NTR}-cRNA (+) and from uninjected oocytes (-) were precipitated with wheat germ lectin-agarose followed by immunoblotting with an antibody against the extracellular domain of p75^{NTR} (REX). C, a chimeric EGFR/p75^{NTR} receptor containing the extracellular domain of the EGF receptor fused to the transmembrane and cytoplasmic domains of p75^{NTR} does not inhibit NT-3 signaling through TrkA. Oocytes expressing TrkA alone or in combination with EGFR/p75^{NTR} were loaded with ⁴⁵Ca²⁺. After stabilization of ⁴⁵Ca²⁺ efflux levels, 200 ng/ml NT-3 was added to the medium (indicated by arrow), and calcium efflux was measured every 10 min. D, oocytes injected with the EGFR/p75^{NTR} cRNA express the chimeric receptor protein. Lysates from oocytes injected with EGFR/p75^{NTR} cRNA (+) and from uninjected oocytes (-) were immunoprecipitated with monoclonal antibody that recognizes the extracellular domain of the EGF receptor followed by immunoblotting using the anti-EGF receptor antibody.

μg/ml of BDNF, or with 50 μg/ml of anti-p75^{NTR} antibody (REX), or in medium alone. BDNF and anti-p75^{NTR} antibody at these concentrations nearly abolished NT-3 binding to p75^{NTR}

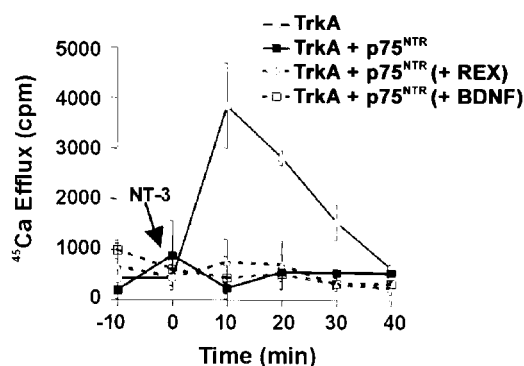


FIG. 5. p75^{NTR} does not need to directly bind to NT-3 to inhibit activation of signaling through TrkA. Oocytes expressing TrkA alone or in combination with p75^{NTR} were loaded with ⁴⁵Ca²⁺. Oocytes expressing TrkA and p75^{NTR} were treated for 2 h with 50 μg/ml of anti-p75^{NTR} antibody (REX), 1 μg/ml BDNF, or were left untreated as indicated. The anti-p75^{NTR} antibody and BDNF at this concentration have both been shown to prevent NT-3 binding to p75^{NTR} (see "Results"). After stabilization of ⁴⁵Ca²⁺ efflux, 200 ng/ml of NT-3 was added at time 0 (indicated by arrow), and calcium efflux was measured every 10 min.

in PC12 cells overexpressing TrkA (data not shown), and the ability of the anti-p75^{NTR} antibody to block NT-3 binding to p75^{NTR} in oocytes was verified by binding and cross-linking studies (data not shown). 200 ng/ml of NT-3 was added to the oocytes, and ⁴⁵Ca²⁺ efflux was measured every 10 min. The data show that blocking NT-3 binding to p75^{NTR} had no effect on the ability of this receptor to block TrkA activation (Fig. 5). No response to NT-3 was seen in the presence of the p75^{NTR} antibody or BDNF. This experiment shows that p75^{NTR} inhibited NT-3 signaling through TrkA, even in the absence of direct binding to NT-3. Because BDNF and the anti-p75^{NTR} antibody may act as p75^{NTR} ligands, this experiment does not exclude the possibility that p75^{NTR} liganding may alter TrkA responsiveness to NT-3. However, it does indicate that direct interaction of p75^{NTR} with NT-3 is not required to inhibit TrkA activation.

p75^{NTR} and TrkA Physically Interacted in *Xenopus* Oocytes Coexpressing Both Receptors—Our results suggested that TrkA and p75^{NTR} may physically interact in surface membranes. To address this possibility, we coexpressed a C-terminal FLAG-tagged TrkA receptor along with p75^{NTR} in *Xenopus* oocytes. The TrkA construct consisted of full-length TrkA with two FLAG epitopes in tandem at the C terminus. Expression of the FLAG-tagged receptor was verified by homogenizing 15 oocytes microinjected with cRNA encoding the receptor, immunoprecipitating with a monoclonal antibody against the FLAG epitope (M2), and immunoblotting with the same anti-FLAG antibody. A specific doublet corresponding to TrkA was identified, which was not present in uninjected oocytes (Fig. 6).

We next asked if the FLAG-tagged TrkA receptor functionally responded to neurotrophin stimulation. NGF activation of TrkA results in activation of the phospholipase C-γ pathway with resultant intracellular calcium release (39). Because a calcium-activated chloride current occurs in *Xenopus* oocytes in response to release of intracellular calcium (32, 33, 35), we performed electrophysiologic recordings from single oocytes in the presence of NGF to confirm that FLAG-tagged TrkA was functional. Oocytes expressing this receptor responded specifically to 100 ng/ml of NGF,² demonstrating that this receptor is indeed functional.

To determine whether p75^{NTR} physically interacted with

² P. S. Mischel and J. Umbach, S. Eskandari, S. G. Smith, C. B. Gundersen, and G. A. Zampighi, submitted for publication.

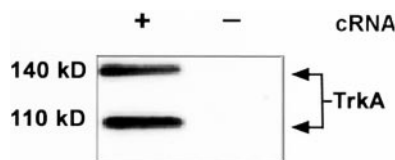


FIG. 6. Expression of FLAG-tagged TrkA receptor in microinjected *Xenopus* oocytes. Fifteen oocytes were injected with 1 ng of TrkA-FLAG cRNA (+) or were left uninjected (-). Thirty six hours after injection, oocytes were lysed, immunoprecipitated with anti-FLAG antibody, separated by 7.5% SDS-PAGE, and immunoblotted with the anti-FLAG antibody (see "Experimental Procedures"). Bands of 140 and 110 kDa are seen in the TrkA-injected oocytes.

TrkA, we performed coprecipitation studies in microinjected oocytes. Equal amounts of cRNA for FLAG-tagged TrkA and p75^{NTR} (5 ng of each cRNA) were injected, and after 48 h, oocytes were exposed to NT-3 (100 ng/ml), NGF (100 ng/ml), or no neurotrophin for 20 min at room temperature, after which they were placed on ice (to prevent further membrane trafficking events).

Oocytes were manually homogenized and fractionated into a heavy fraction, containing the membrane and cortical granules, and a cytoplasmic fraction (see "Experimental Procedures"). Fractions were solubilized in a buffer containing 2% Triton X-100 (and protease inhibitors) for 15 min at 37 °C and were then centrifuged. Supernatants were pre-cleared with protein A/G-Sepharose beads and immunoprecipitated with a polyclonal antibody to p75^{NTR} (25). SDS-PAGE was performed on samples (see "Experimental Procedures"). After transfer to nitrocellulose membranes, samples were immunoblotted with the monoclonal antibody to the FLAG epitope (Stratagene, La Jolla, CA). As seen in Fig. 7, TrkA was coprecipitated with p75^{NTR} from the membrane fractions of oocytes injected with TrkA and p75^{NTR} (lanes 2–4) but not from uninjected oocytes (lane 5) or from the antibody plus protein A/G bead control sample (lane 1). p75^{NTR} and TrkA were coprecipitated in oocytes expressing these receptors regardless of whether they were treated with 100 ng/ml of NT-3 (lane 4), NGF (lane 3), or with no neurotrophin (lane 2). The anti-p75^{NTR} serum has previously been shown to bind p75^{NTR} and to lack detectable binding to TrkA (25). We further verified that the anti-p75^{NTR} serum did not immunoprecipitate TrkA from oocytes expressing TrkA alone and that the anti-FLAG monoclonal antibody did not recognize a specific band in oocytes expressing only p75^{NTR} (data not shown). Because high levels of expression of both p75^{NTR} and TrkA were required to observe this interaction, and only a relatively small percentage of each receptor appears to be in this complex, we cannot fully determine the role of this physical interaction at physiological concentrations. However, these results do suggest that p75^{NTR} and TrkA can physically interact in the plasma membrane and that this interaction appears to be independent of the presence of neurotrophins.

DISCUSSION

In this paper, we examined the role of p75^{NTR} in inhibiting NT-3 signaling through TrkA. By using a calcium efflux assay to assess signaling in *Xenopus* oocytes microinjected to express both receptors, we demonstrated quantitatively that p75^{NTR} completely suppressed NT-3 signaling through TrkA. The inhibition was specific to NT-3, because NGF signaled equally well through TrkA, regardless of whether or not p75^{NTR} was present. Our data are consistent with studies in PC12 cells which demonstrate that p75^{NTR} suppresses NT-3 signaling through TrkA (4, 23) and with the observation that TrkA-expressing sympathetic neurons from p75^{NTR}−/− mice are more responsive to NT-3 than wild type sympathetic neurons

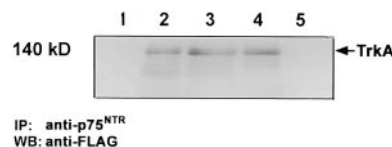


FIG. 7. p75^{NTR} and TrkA can be coimmunoprecipitated from the membrane fractions of microinjected *Xenopus* oocytes expressing both receptors. Oocytes were injected with TrkA-FLAG and p75^{NTR} (5 ng each). Thirty six hours later, 12 oocytes per condition were treated with 100 ng/ml NGF (lane 3), 100 ng/ml NT-3 (lane 4), or no neurotrophin (lane 2) for 20 min at room temperature (see "Experimental Procedures"). Oocytes were lysed, immunoprecipitated with anti-p75^{NTR} antibody, and immunoblotted with anti-FLAG antibody. TrkA is coprecipitated with p75^{NTR} in oocytes injected with TrkA and p75^{NTR} cRNA (lanes 2–4), but not in uninjected oocytes (lane 5), or in antibody and protein A/G-Sepharose beads only control (lane 1). Anti-p75^{NTR} serum has previously been shown to bind p75^{NTR} and to lack detectable binding to TrkA (25). We have also observed that anti-p75^{NTR} anti-serum does not precipitate TrkA from oocytes expressing TrkA only, nor does the anti-FLAG monoclonal antibody recognize a specific band from oocytes expressing p75^{NTR} alone (data not shown).

(24). Our findings are also consistent with the recent evidence that p75^{NTR} can inhibit the ability of NT-3 to activate TrkA *in vivo* (40). Compared with control littermates, NGF+/− mice have a 50% deficit in sympathetic neurons. This deficit is rescued by NT-3 in the absence of p75^{NTR} (NGF+/− and p75^{NTR}−/−) but not in mice that express normal levels of p75^{NTR} (40). The rescue of these TrkA-expressing sympathetic neurons depends on NT-3, since many of these neurons are lost in NGF+/−, p75^{NTR}−/−, NT-3+/− mice (40).

In addition to inhibiting TrkA signaling, p75^{NTR} may also inhibit signaling through TrkB. In transfected A293 cells expressing TrkB, coexpression of p75^{NTR} decreased TrkB phosphorylation in response to NT-3 and NT-4 but not in response to BDNF (41). In contrast to this finding, another group (42) has recently demonstrated that coexpression of p75^{NTR} with TrkB inhibited phosphorylation in response to BDNF and NT-4 but not NT-3. The full effect of p75^{NTR} on TrkB signaling in response to BDNF, NT-4, and NT-3 signaling through TrkB will need to be further elucidated. However, the mechanism by which p75^{NTR} suppresses TrkB activation may be similar to the way p75^{NTR} suppresses NT-3 signaling through TrkA.

Structural Domains of p75^{NTR} Involved in the Inhibition of NT-3 Signaling through TrkA—In these experiments, we showed that the extracellular domain of p75^{NTR} was necessary to inhibit NT-3 signaling through TrkA. NT-3 could not signal through TrkA in *Xenopus* oocytes coexpressing a mutant p75^{NTR} receptor that lacks all but the first nine amino acids of its cytoplasmic domain. In contrast, oocytes coexpressing an EGFR/p75^{NTR} chimeric receptor, which contained full p75^{NTR} transmembrane and cytoplasmic domains, did not inhibit NT-3 signaling through TrkA. Based on these findings, we conclude that the extracellular domain of p75^{NTR} is necessary to inhibit NT-3 signaling through TrkA. These experiments also clearly showed that the transmembrane and cytoplasmic domains of p75^{NTR} are not sufficient to inhibit NT-3 signaling through TrkA.

The truncated p75^{NTR} receptor contains the juxtamembrane region of p75^{NTR}, including cysteine 279, which is critical for palmitoylation of p75^{NTR} (43). Because palmitoylation of p75^{NTR} appears to be important for targeting p75^{NTR} (as well as TrkA) to caveolae-like membranes where much of TrkA receptor signaling is initiated (44), it is possible that the juxtamembrane region of p75^{NTR} may be important for inhibiting NT-3 signaling through TrkA. However, our experiments clearly show that this region, in the absence of the extracellular domain, is not sufficient to inhibit NT-3 signaling through TrkA.

p75^{NTR} and Trk receptors may engage in cross-talk through the interaction of their cytoplasmic signaling pathways. To date, much of this interaction appears to be inhibitory. NGF activation of TrkA can block the p75^{NTR}-mediated cell death pathway (21), and activation of p75^{NTR} by BDNF suppresses cellular responses to NGF activation of TrkA (37, 38). Our data do not entirely exclude the possibility that p75^{NTR} initiates a cytoplasmic signal through its juxtamembrane region, either directly or by interacting with another cytoplasmic protein. However, our observation that the p75^{NTR} receptor, lacking almost the entire cytoplasmic domain, inhibited NT-3 signaling through TrkA suggests that p75^{NTR} did not act downstream from the receptor to suppress TrkA signaling.

Interaction of p75^{NTR} and TrkA to Mediate the Inhibition of NT-3 Signaling through TrkA—By coexpressing TrkA and p75^{NTR} in oocytes and occupying p75^{NTR} with either BDNF or anti-p75^{NTR} antibody, we determined that p75^{NTR} inhibited activation of TrkA even in the absence of direct binding to NT-3. Because BDNF and REX may act as p75^{NTR} ligands, these results do not exclude the possibility that liganding of p75^{NTR} alters the ability of TrkA to respond to NT-3. More importantly, these results suggest strongly that events occurring between p75^{NTR} and TrkA, either directly or indirectly, are critical for inhibiting TrkA response to NT-3.

In oocytes coexpressing both receptors at significantly higher levels than used in the ⁴⁵Ca²⁺ efflux assays, we coimmunoprecipitated p75^{NTR} and TrkA from the membrane fraction of oocytes. Although we could only detect this interaction when both receptors were expressed at these relatively higher levels, these findings suggest that p75^{NTR} and TrkA can physically interact in the surface membrane and that this interaction does not require the presence of neurotrophins.

The structural basis for p75^{NTR} and TrkA interactions has not been defined. Evidence to support their interaction has been presented (41, 45–49), and most reports have focused on the role of p75^{NTR} in increasing the affinity of TrkA for NGF (19, 48, 49). Our results suggest a potential role for the physical interaction of p75^{NTR} and TrkA in regulating the specificity of TrkA for NGF relative to NT-3. The coimmunoprecipitation data suggest the existence of either a direct physical interaction between p75^{NTR} and TrkA or the formation of a complex bridged by one or more mutually interacting proteins. p75^{NTR} and TrkA are both significantly enriched in caveolae-like domains at the plasma membrane, and the two receptors may interact (at least functionally) within this micro-environment (44). Furthermore, indirect interaction between p75^{NTR} and TrkA via the mutually interacting protein caveolin-1 has been suggested (50).

It is also possible that p75^{NTR} inhibits NT-3 signaling by inhibiting NT-3 binding to TrkA. However, preliminary data from our laboratory using ¹²⁵I-NT-3 binding and cross-linking in PC12 cells that overexpress TrkA indicate that NT-3 can still bind TrkA in the presence of p75^{NTR}.³ In these cells, occupying p75^{NTR} with BDNF or REX did not alter the amount of NT-3 bound to TrkA.³ However, since BDNF and REX may act as p75^{NTR} ligands, it is possible that liganded p75^{NTR} alters the conformation of the TrkA-binding site. An allosteric model for p75-*TrkA* interactions has been proposed that may account for this property (51). In the future, it will be important to examine NT-3 binding in a cell line that expresses TrkA alone, or in combination with p75^{NTR}, to fully determine the effect of p75^{NTR} on NT-3 binding to TrkA.

In summary, our experiments have shown that p75^{NTR} in-

teracts with TrkA to inhibit NT-3 signaling. Our work demonstrates that the extracellular domain of p75^{NTR} is necessary for this inhibition and suggests that this interaction occurs at the level of p75^{NTR}-TrkA interaction on the surface membrane. Our study has important implications for understanding the regulation of NT-3 signaling through TrkA *in vivo*. During late embryonic and early postnatal development, nearly 50% of sympathetic neurons that are NGF-dependent develop an additional requirement for NT-3, mediated by TrkA (8, 9, 52, 53). These neurons express low levels of p75^{NTR} relative to TrkA (53), which our data suggest would enable NT-3 to promote survival by activating TrkA (2, 5, 53). As targets are reached, p75^{NTR} expression increases and responsiveness to NT-3 declines (2). *In vivo* studies of the neonatal sympathetic ganglia show that p75^{NTR} regulates the survival of TrkA-expressing sympathetic neurons by regulating the responsiveness of these neurons to NT-3 (40). In fact, it appears that one of the most important roles for p75^{NTR} in the developing sympathetic ganglia is to regulate TrkA responsiveness to NT-3 (40). In addition to sympathetic neurons, a subset of late embryonic TrkA-expressing sensory neurons that are NGF-dependent also develop a requirement for NT-3 that is mediated by TrkA (5, 10). Unlike postnatal sympathetic neurons, postnatal sensory neurons lose their ability to respond to NT-3 via TrkA. It will be interesting to determine whether p75^{NTR} plays a role in decreasing the responsiveness of postnatal sensory neurons to NT-3. Finally, although our work has focused on the role of p75^{NTR} in regulating NT-3 signaling through TrkA, p75^{NTR} may interact with TrkB to regulate its responsiveness. It will be important in the future to determine whether p75^{NTR} regulates TrkB signaling by a similar mechanism.

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